



750xg, 2 mins



750xg, 3 mins



Purified Sample



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## **HiYield™ G50 Cleanup Kit** **Protocol Book**

*Better Sequencing Data*

Cat. No. **YCG50**

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**Real Genomics™**

## **HiYield™ G50 Cleanup Kit**

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## HiYield™ G50 Cleanup Kit

Cat. No. **YCG50**  
50 preps / kit  
G50 Column: 50 pcs  
2ml Collection Tube: 50 pcs



**Sample:** 20-50 µl  
**Recovery:** 90%  
**DNA Size:** Larger than 20 bases  
**Format:** Spin Columns  
**Operation:** Centrifuge  
**Operation Time:** 5 Minutes

### Description

HiYield™ G50 Cleanup Kit consists of prepacked Sephadax G50 pre-hydrated with double-distilled water. This kit is ideal for removing excess dye terminator, free nucleotides from sequencing and labeling reaction, desalting and for buffer exchange. G50 column can purify DNA fragments larger than 20 bases in length with low molecular weight material retained in the gel matrix of the column. This kit is designed to purify DNA fragments larger than 20 bp only. Not suitable for PCR product primer removal.

### Features

Entire procedure could be completed within 5 minutes.  
Ready-to-use prehydrated gel-filtration columns.  
Optimized for efficient removal of any dye terminator.

### Applications

Desalting, Dye Terminator Removal, Buffer Exchange.

### Quality Control

The quality of HiYield™ G50 Cleanup Kit is tested on a lot to lot basis. The particle size and quality is tested. The purified DNA is checked by electrophoresis.

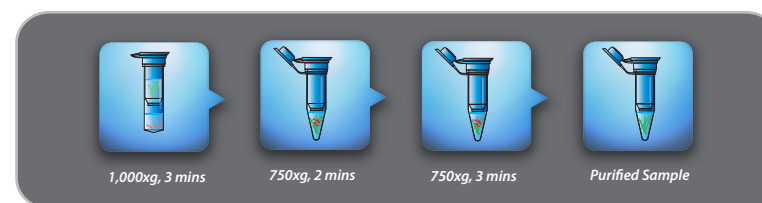
*Note: For research use only. This kit contains irritant agent. During operation, always wear a lab coat, disposable gloves and protective goggles.*

### Purification / Desalting Protocol



1. Place a G-50 Column in a 2 ml Collection Tube.
2. Centrifuge at 750 x g for 2 minutes.
3. Transfer the G-50 Column to a 1.5 ml microcentrifuge tube.
4. Carefully load the sample (20-50 µl) onto the center of the gel bed surface.
5. Centrifuge again at 750 x g for 3 minutes. The purified sample can be recovered at the bottom of the 1.5 ml microcentrifuge tube (approximately the same volume as the loaded sample).

### Buffer Exchange Protocol



1. Place a G-50 Column in a 2 ml Collection Tube.
2. Centrifuge at 1,000 x g for 3 minutes.
3. Discard the flow-through in the 2 ml Collection Tube and place the G-50 Column back in the same 2 ml Collection Tube.
4. Add 350 µl of desired buffer to the G-50 Column. Then Centrifuge at 750 x g for 2 minutes.
5. Transfer the G-50 Column to a 1.5 ml microcentrifuge tube.
6. Carefully load the sample (20-50 µl) onto the center of the gel bed surface.
7. Centrifuge again at 750 x g for 3 minutes. The purified sample can be recovered at the bottom of the 1.5 ml microcentrifuge tube (approximately the same volume as the loaded sample).